

Exploring fungal diversity and interactions with *L. acicola* in Brown Spot Needle Blight

**Emmanuel D. Nyarko, Annakay Newell,
Rabiu Olatinwo and Lori G. Eckhardt**

Forest Health Dynamics Laboratory
College of Forestry, Wildlife and Environment
Auburn University, Auburn AL

Forest Health Dynamics Laboratory

College of Forestry, Wildlife and Environment, Auburn University



Background

- Loblolly pine (*Pinus taeda*) is the most abundant pine found in Alabama
- Widely planted for commercial use
- Thrives in various soil types and climatic conditions, making it ideal for reforestation and afforestation



Problem

Brown spot needle blight poses a significant threat to the productivity and economic viability of loblolly pine plantations

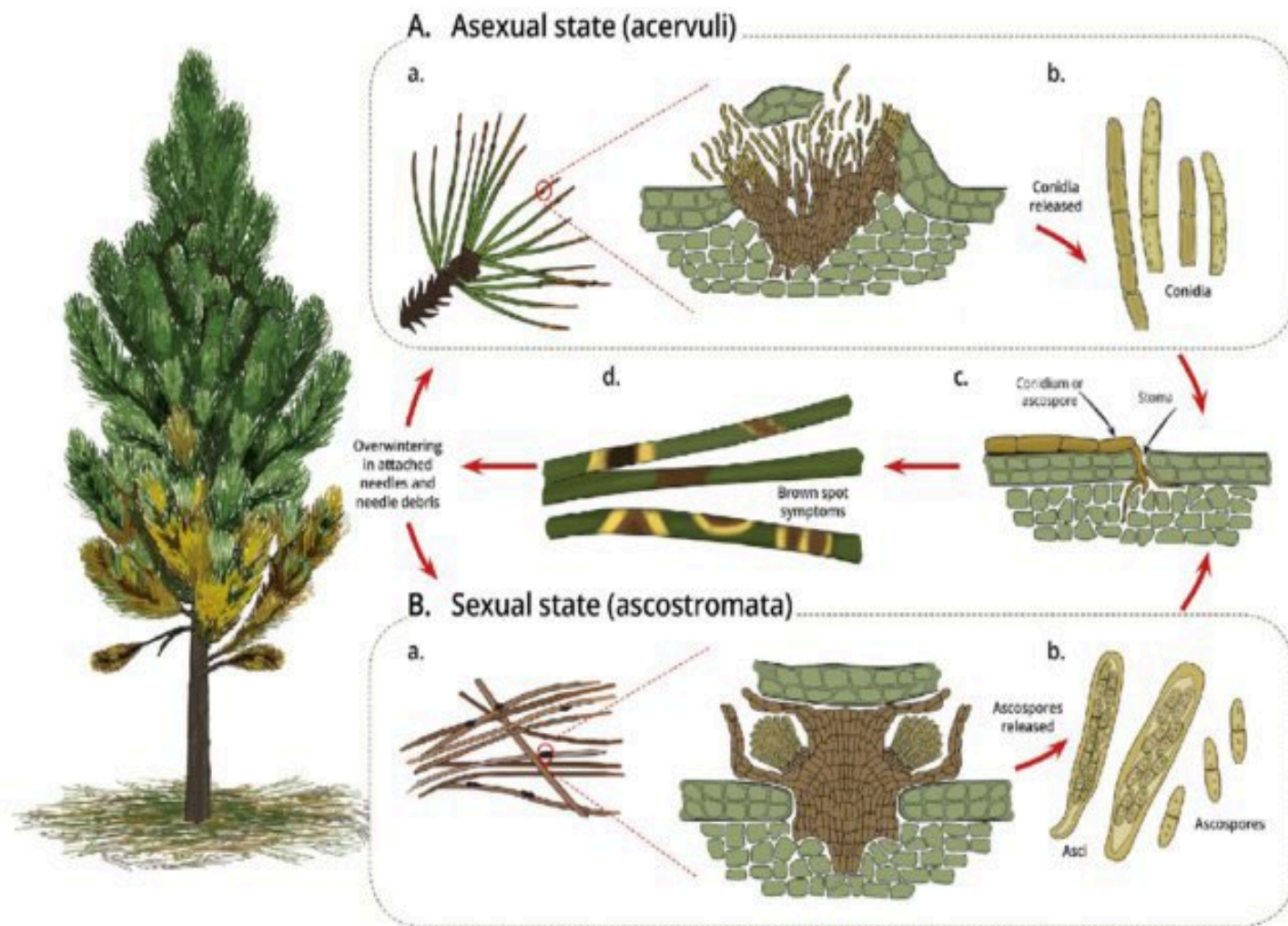
Seedlings in nurseries and young plantations are also vulnerable

BSNB is caused by the fungus, *Lecanosticta acicola*



BSNB infected study plot in Chatom, AL

Life cycle of *L. acicola*



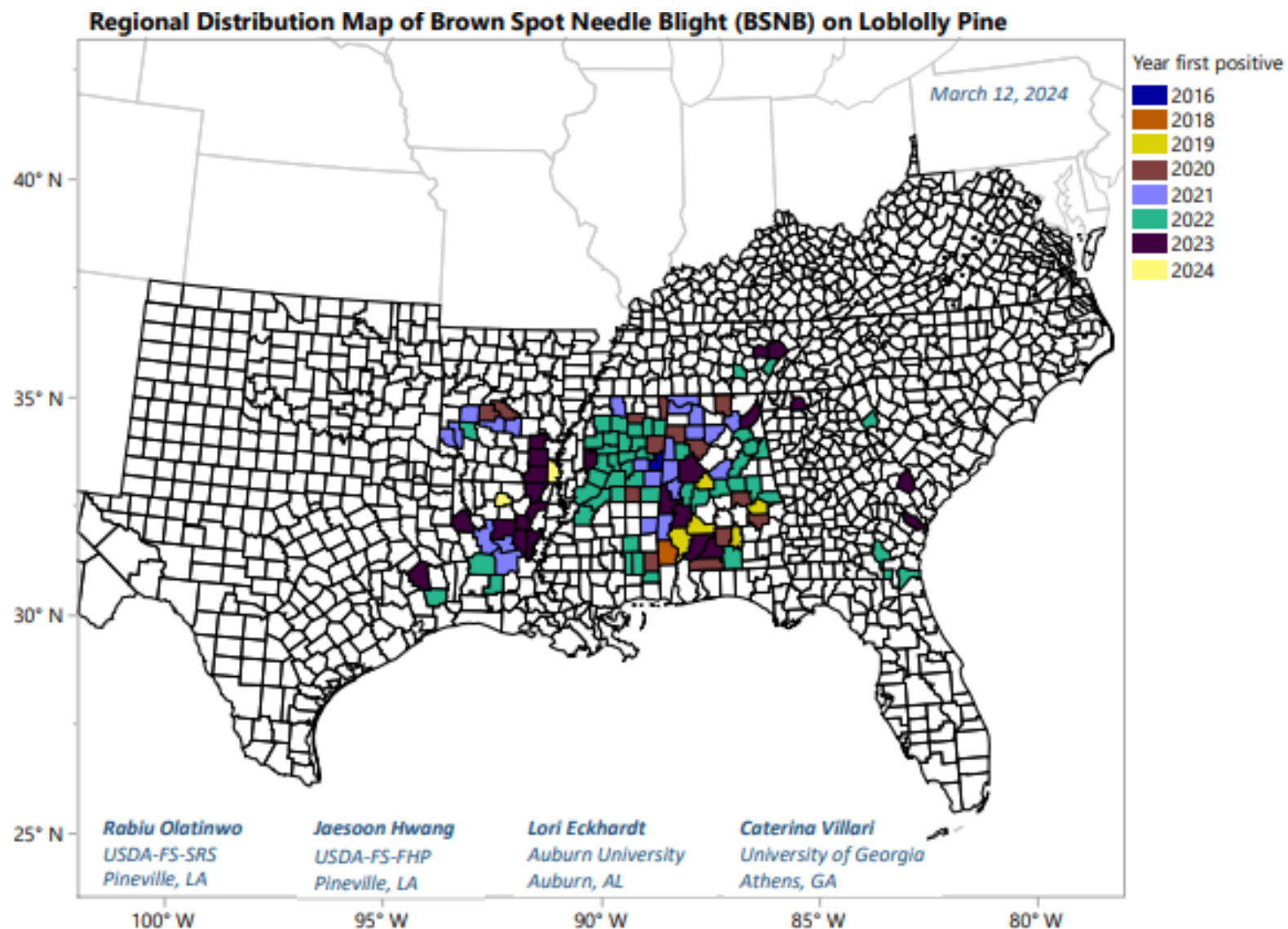
Van Der Nest et al 2019

Impact of Brown Spot Needle Blight



Datta, D. & Eckhardt L.G. 2021

- Reduced growth
- Premature shedding of needles
- Tree mortality



Contact: Rabiu Olatinwo (rabiu.o.olatinwo@usda.gov)

Objectives

Study 1

To investigate the diversity of fungi associated with *Lecanosticta acicola* in brown spot needle blight

Study 2

To assess the seedling susceptibility of different loblolly pine families to *Lecanosticta acicola*

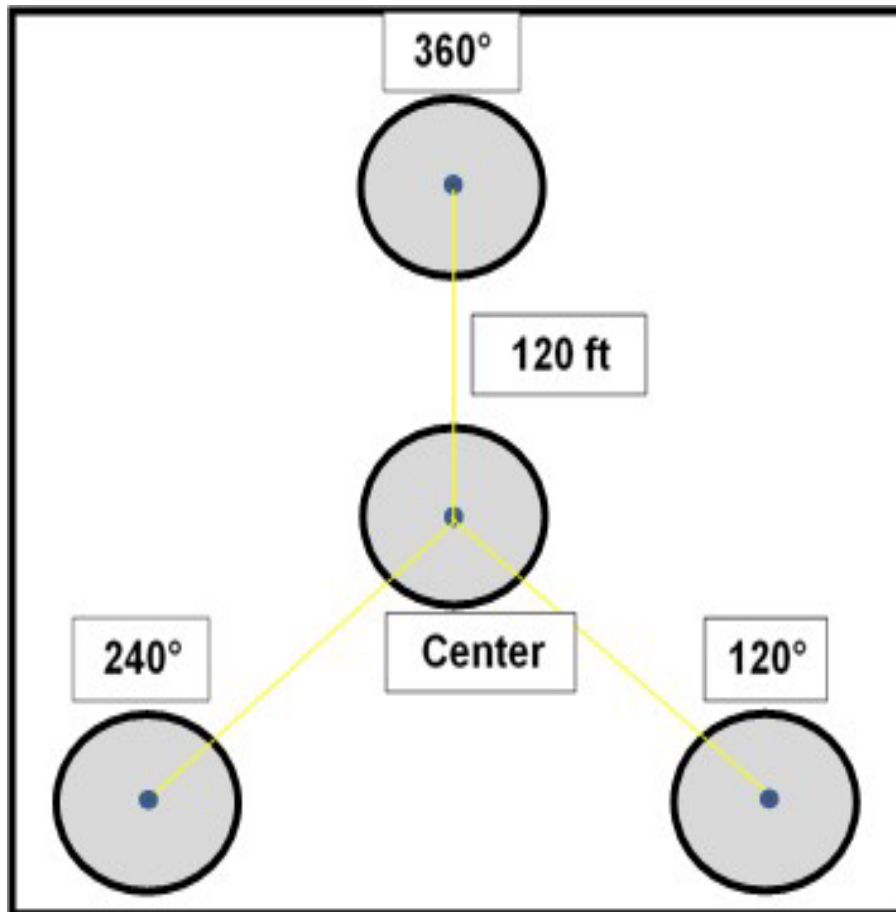
Study Areas

Research plots located in Cullman and Chatom,
Alabama

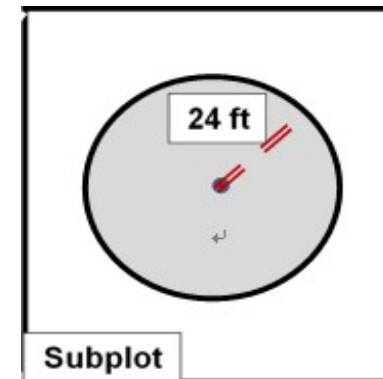
14 plots in 2023

3 National Forests (8 new plots)- January 2024
Bankhead NF, Conecuh NF, Tuskegee NF

Plot layout



Area: ≈ 2.2 acres



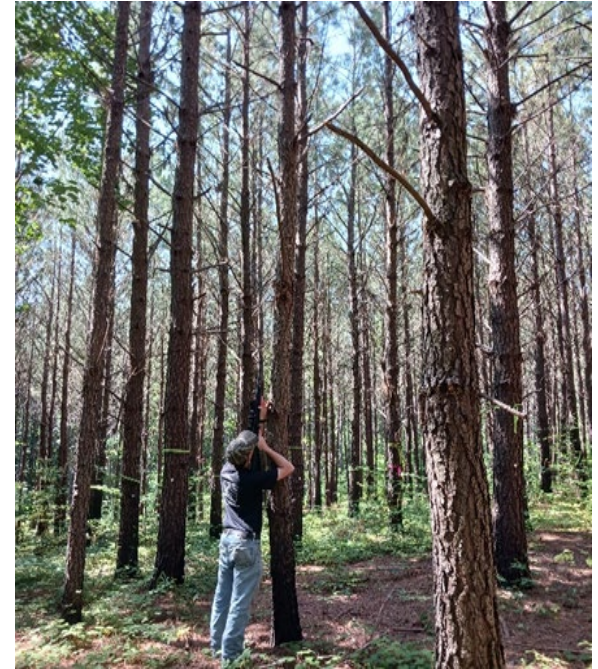
FIA Plot design for the Forest Health Monitoring Program, 1990

Needle collection

Sampling period – monthly
(March to November)

Needle samples collected from one
tree per subplot in all plots

Transported to the Forest Health
Lab for processing



Processing and Identification

1

Needle Examination

**Incubated Needles
(Sporulation Chambers)**

2

Needle Examination

Surface Sterilization

**Plated onto Isolation
Media**

**Incubation (20-25°C
for 5 days to 2 Weeks)**

**Transfer to
Identification Media**

**Morphological
Identification**

3

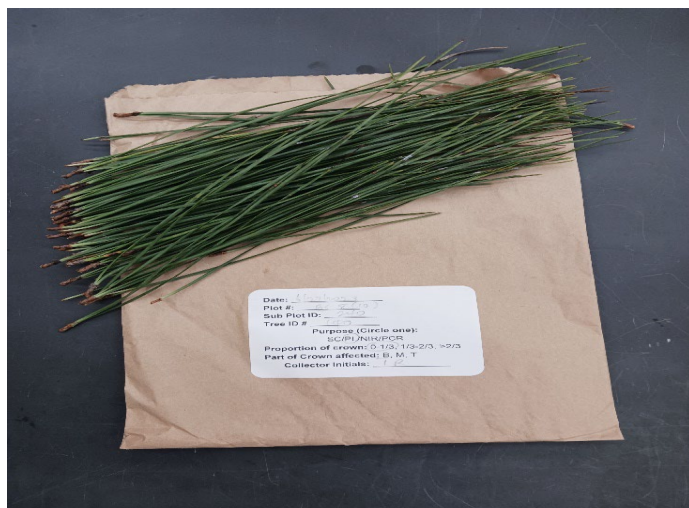
Needle Examination

DNA Extraction

PCR

**Molecular
Identification**

Sample processing (Plating)



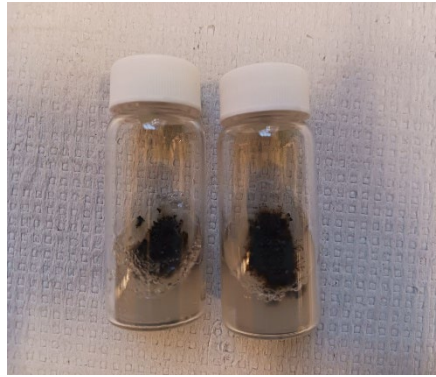
Results

Predominant fungi recovered from plated needles

Lecanosticta acicola



FHDL



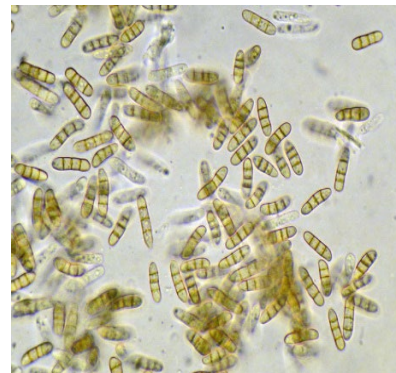
FHDL



Hendersonia



FHDL

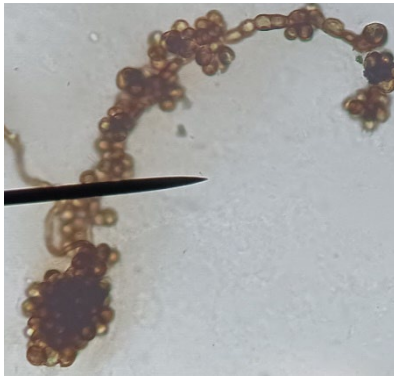


Barnett, H. L., & Hunter B.B., 1998

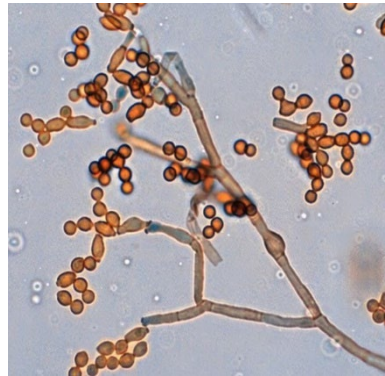
Results

Predominant fungi recovered from plated needles

Cladosporium



FHDL

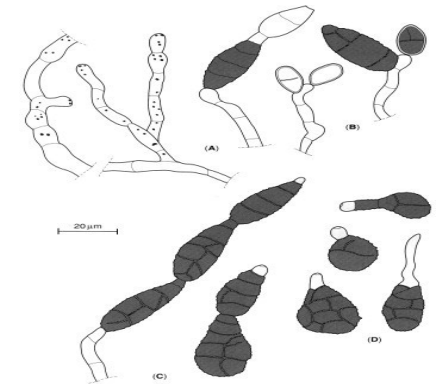


Barnett, H. L., & Hunter B.B., 1998

Alternaria

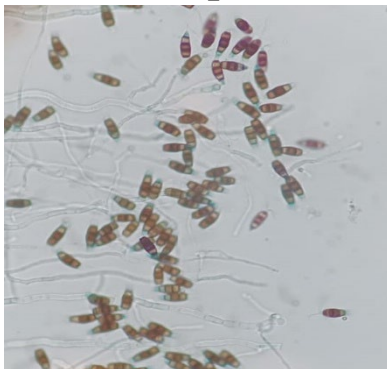


FHDL

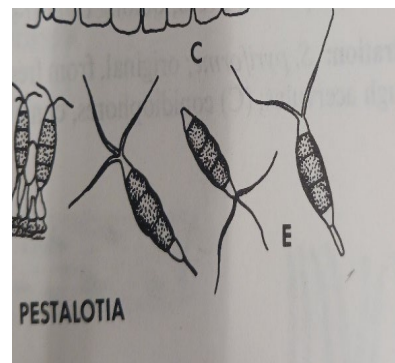


Barnett, H. L., & Hunter B.B., 1998

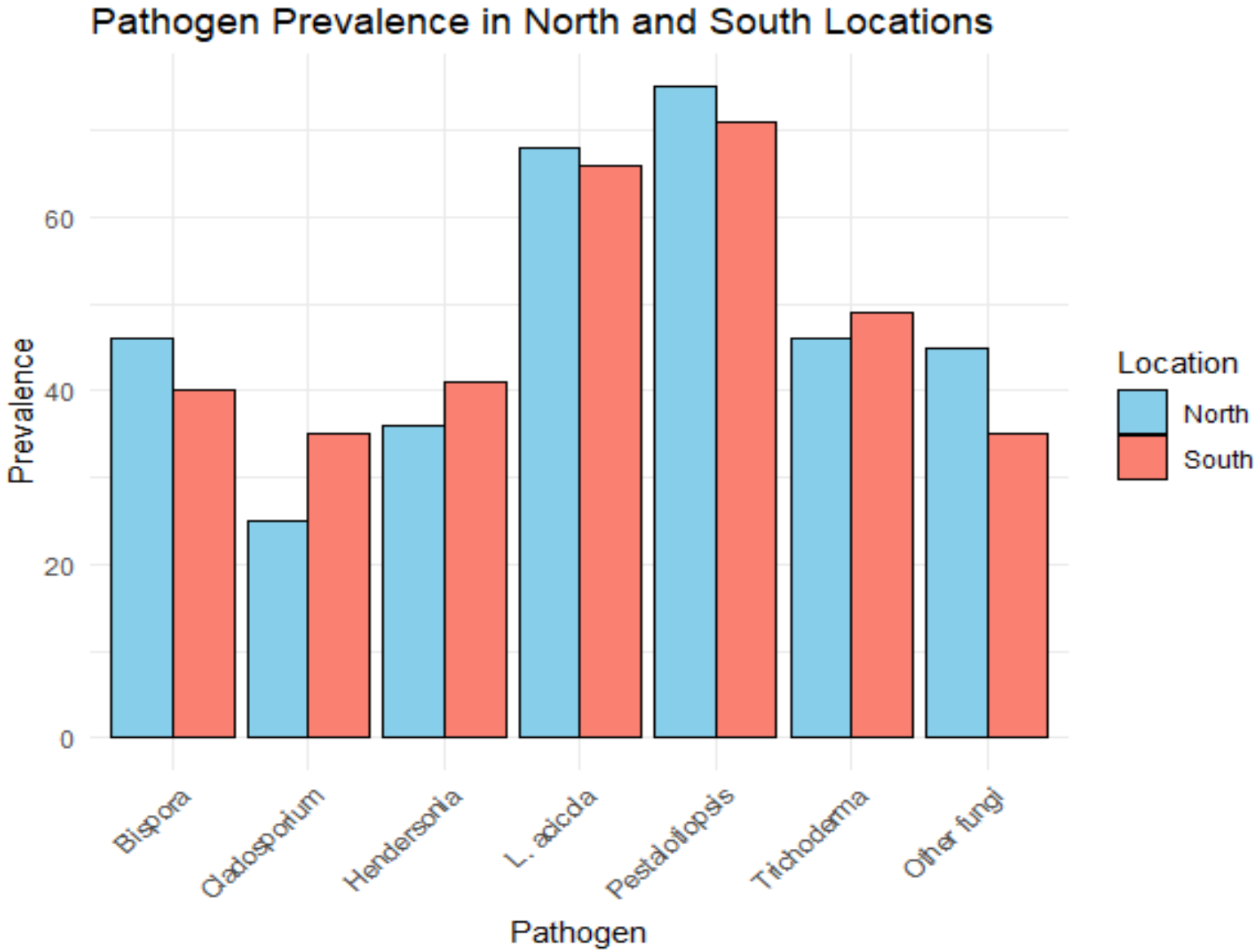
Pestalotiopsis

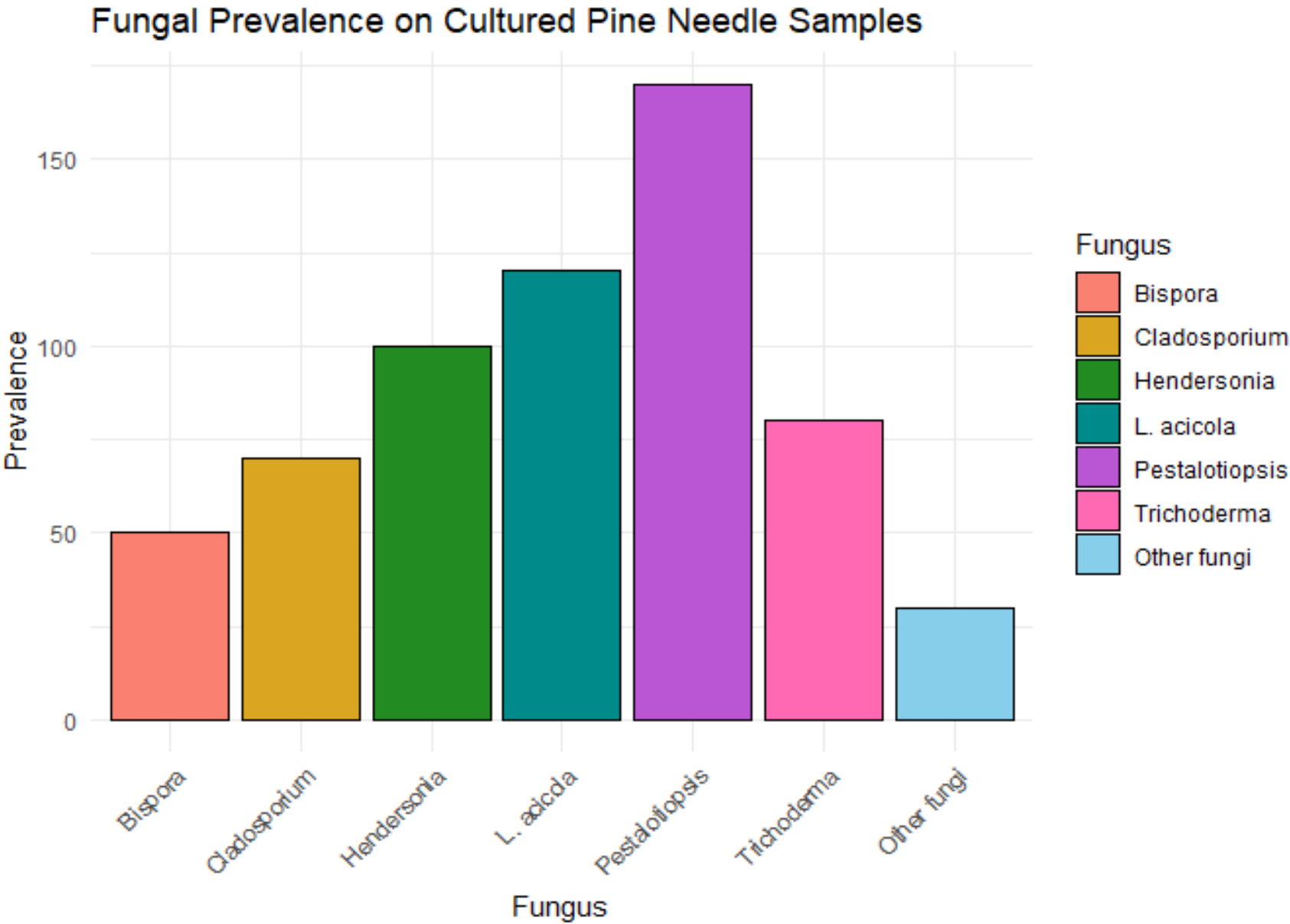


FHDL



Barnett, H. L., & Hunter B.B., 1998





Ecological roles of predominant fungi

| Fungi | Ecological role |
|-----------------------|-------------------------------|
| <i>L. acicola</i> | Parasitic |
| <i>Pestalotiopsis</i> | Endophytic, saprophytic |
| <i>Cladosporium</i> | Weakly parasitic, saprophytic |
| <i>Alternaria</i> | Parasitic, Saprophytic |
| <i>Trichoderma</i> | Saprophytic |
| <i>Bispora</i> | Saprophytic |

Barnett, H. L., & Hunter B.B., 1998. Illustrated Genera of Imperfect Fungi 4th Edition

On-going work

- DNA extraction for samples collected in 2023
- Needle sample collection on newly added National Forests
- Seedling susceptibility experiment (Study 2)

Barnett, H. L., & Hunter B.B., 1998. Illustrated Genera of Imperfect Fungi 4th Edition

Acknowledgment

Dr. Lori Eckhardt (Advisor)

Committee Members

FHDL Members (Graduate and undergraduate students)

Kris Bradley

Dr. Glenn Glover & Family (Osko Forest)

The Williams Family

Stallworth Land Company

Longleaf Land & Timber Company



AUBURN
College of Forestry,
Wildlife and Environment



**Forest Health
Cooperative**



THANK YOU